

Yeast DNA Prep for PCR (Smash and Grab)
(Madhani Lab)

Place 200 ul of Smash and Grab buffer in Eppie tube.

Add cells (e.g. a toothpick-full)

Add glass beads (50-200 ul)

Add 200 ul equilibrated phenol

Cap and vortex (use a platform vortexer) for 2-5 min.

Add 200 ul water

Spin in a microfuge at top speed for 5 min.

Take 1 ul of supernatant and add to 100 ul of water.

Use 1 ul in a 50 ul PCR.

-or-

for a cleaner prep (e.g. for plamid rescue):

Transfer supernatant (after 5min spin) to a new tube and add 1 ml of ethanol, mix.

Spin 5 min at top speed.

Wash pellet with 500 ul 100% ethanol, dry in speedvac.

Resuspend in 50 ul of TE or water

S&G buffer (aka breaking buffer from the Yeast section of Current Protocols)

2% (v/v) Triton X-100

1% (v/v) sodium dodecyl sulfate (SDS)

100 mM NaCl

10 mM Tris·Cl, pH 8.0

1 mM EDTA, pH 8.0