

# **Denatured yeast protein extracts by NaOH lysis/TCA precipitation**

according to Knop et al. 1999, *Yeast* 15(10B):963–72

modified by Sigurd Braun

## use:

quick preparation of protein samples from *S. cerevisiae*, *S. pombe*, *C. neoformans*  
(avoidance of glass bead disruption/phenol/boiling)

time: less than 60 minutes

## **Buffers/Solution:**

- **NaOH/BME:** 138.75  $\mu$ l 2N NaOH + 11.25  $\mu$ l BME per sample  
(usually, for 12 samples a mix of 1850  $\mu$ l NaOH + 150  $\mu$ l BME is prepared)
- **TCA:** 55% (w/v) trichloroacetic acid in H<sub>2</sub>O (stored at 4 deg)
- **HU buffer:** 200 mM phosphate buffer, pH 6.8, 8 M urea, 5% w/v SDS, 1 mM EDTA, 100 mM DTT, bromophenol blue; stored at –20 deg w/o DTT; DTT is added freshly from 1 M stock)

## **cell lysis and protein precipitation:**

1. harvest yeast cells corresponding to an OD<sub>600</sub> = 1-2
2. resuspend cell pellet in 1 ml ice-cold water
3. add 150  $\mu$ l NaOH/BME to each sample
4. incubate on ice with occasional vortexing for 15 min
5. add 150  $\mu$ l TCA to each sample
6. incubate on ice with occasional vortexing for 15
7. spin down in a table centrifuge at 4 deg at max. speed (e.g. 20K) for 20 min
8. take off most of supernatant with pipette or with vacuum pump tip/syringe needle
9. spin again briefly at max. speed for 1 min
10. carefully take off remaining supernatant with precaution  
(protein pellet may be loose)
11. resuspend pellet in 50  $\mu$ l HU buffer
12. optional: when buffer turns yellow due to some residual TCA in the resolved protein pellet, add some (10-20)  $\mu$ l of 1 M Tris pH 6.8

alternatively, after taking off supernatant (step 4), add 100  $\mu$ l of chilled acetone (–20 deg) to remove any residual TCA; continue with step 5  
(only recommended when protein amounts are precipitated corresponding to more than 1 OD<sub>600</sub> of yeast cells)

## **sample denaturation/SDS PAGE loading:**

1. heat denature at 65-70 deg for 10-15 min  
(do NOT boil as urea denatures/aggregates at higher temperatures than 70 deg)
2. spin briefly in table centrifuge (RT, 12K, 1 min)
3. load 5-10  $\mu$ l of sample (corresponds to 0.1 – 0.2 OD<sub>600</sub> of yeast cells)