

# Plating of yeast cells using a pin-frogger (serial dilutions)

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## Use:

Quick and uniform plating of serial dilutions of yeast cultures  
(better results than using multi-channel pipette or grid pattern)

## Requirements:

empty Petri dish with EtOH; agar (e.g. YPD plate); sterile 96 well microtiter plate; 48 pin or 96 pin frogger w/ round pins of 4 mm diameter

1. dilute mid-log phase grown yeast cultures (in YPD or selective media) to  $OD_{600} = 0.1$  (equals  $1.5-2 \times 10^6$  cells per ml) with sterile water (alternatively, a stationary culture may also be used)
2. transfer 250  $\mu$ l of each diluted cell suspension to first column of 96-well microtiter plate
3. add into the following 5 columns 200  $\mu$ l sterile water
4. make serial dilutions 1:5 using a multi-channel pipette transferring 50  $\mu$ l from each column to the following
5. plate diluted cell suspension series using a pin frogger (stamp, pin diameter 0.3 cm):
  - a. sterilize pin frogger by EtOH/flame
  - b. place sterilized pin frogger on a YPD (or any other plate) for 1-2 min to let it cool down
  - c. dip pin frogger briefly into the cell suspension wells of the microtiter plate
  - d. lift up frogger quickly to avoid losing cell suspensions drops from the pins
  - e. place frogger carefully on the plate on which dilution series should be analyzed and leave it there for 20-25 seconds to ensure that the drops have been come off efficiently from the pins  
(shorter plating times will result in small and uneven droplets; note: the diameter of the droplet depends also on the humidity of the plate – if results don't look satisfying, try freshly poured plates)
  - f. repeat step c-e for further plates to be analyzed in parallel (sterilizing frogger is not necessary when using the same cultures/wells)
  - g. sterilize frogger by repeating steps a and b

After use, clean the pins by first rinsing/dipping the frogger in water or EtOH, then subsequently by flaming off the EtOH; otherwise when flaming directly remnants of media and cells will be “baked” on the surface of the pins, which is hard to remove.